

A Study of Methods of Preparation of Nanoparticles

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Introduction:

A wide range of compounds, including proteins, polysaccharides and synthetic polymers, may be produced using nanoparticles. The selection of matrix materials is subject to many factors, including: (a) the size of nanoparticles required; (b) drug's inherent properties, such as aqueous solubility and stability; (c) surface properties such as load and permeability; (d) the level of biodegradability, biocompatibility and toxicity; The majority of nanoparticles were produced by the following three methods: (1) premade polymers dispersion; (2) monomer polymerisation; (3) hydrophilic hydrophilic polymers ionic gelation or coacervation. However, the literature for the creation of nanoparticles has also detailed alternative techniques such as supercritical fluid technology 8 or Particle Replication for Non-Wetting Templates (PRICT) 9. The latter has full control over particle size, form and compound, which may set an example for future industrial mass manufacturing of nanoparticles. Dispersion of preformed polymers is a common technique used to prepare biodegradable nanoparticles from poly (lactic acid) (PLA); poly (D,L-glycolide), PLG; poly (D, L-lactide-co-glycolide) (PLGA) and poly (cyanoacrylate) (PCA).

Methods of Preparation of Nonoparticles:

There are many methods may be utilised in many ways. Method of evaporation of solvents: This technique dissolves the polymer in an organic solvent like dichloromethane, chloroform or ethylic acetate, which also acts as a solvent for hydrophobic substances dissolving. In an aqueous solution that contains a surfactant or emulsifying agent, the combination of the polymer and drug solution is then emulsified into water emulsion (o/w). The organic solvent is evaporated by lowering the pressure or constant mixing after the creation of stable emulsions. The kind and amounts of stabilisers, homogenizer velocity and polymer levels were shown to affect the particulate size. A high-speed homogenization or ultrasonics may frequently be used to create tiny particles.

Spontaneous emulsification or solvent diffusion method

The solvent evaporation method is a modified version of the previous approach. So the watermiscible solvent, in conjunction with a trace amount of the water immiscible organic solvent, is used as an oil phase in the process. Because to the spontaneous diffusion of solvents, the two phases contributing to the formation of small particles are interfacially Turbulent during the manufacturing process. When the concentration of the water miscible solvent increases, it is possible to achieve a reduction in the size of the particles. Both solvent evaporation and solvent diffusion methods may be used to produce hydrophobic or hydrophilic medications. A number of w/o/w emulsions must be produced by hydrophilic pharmaceuticals dissolved in the internal aqueous phase of the drug formulation.

Polymerization method

Monomers are polymerized to produce nanoparticles in an aqueous solution in this technique. The drug is included either by dissolving it in the polymerization



solution or by adsorption onto the nanoparticles after the polymerization process is finished. By ultracentrifugation, the nanoparticle suspension is purified to eliminate different stabilisers and surfactants used in polymerization, and the particles are then resuspended in an isotonic surfactant-free medium. This method has been used to create polybutylcyanoacrylate and poly(alkylcyanoacrylate) nanoparticles16. The concentration of surfactants and stabilisers employed determines the production of nanocapsules and their particle size.

Coacervation or ionic gelation method

The creation of nanoparticles utilising biodegradable hydrophilic polymers such as chitosan, gelatin, and sodium alginate has attracted a great deal of interest in the scientific community. For the production of hydrophilic chitosan nanoparticles, Calvo and colleagues developed an ionic gelation method, which they published in Science. In order to do this, the method makes use of two aqueous phases, one of which is chitosan, which is a diblock co-polymer of ethylene oxide or propylene oxide (PEO-PPO), and the other of which is a polyanion sodium tripolyphosphate (polyanion NTP). When the positively charged amino group of chitosan comes into contact with the negatively charged tripolyphosphate, it forms nanometer-sized coacervates, according to this procedure. It is possible to create coacervates when two aqueous phases come into contact with each other electrostatically, whereas ionic gelation is the transition of a material from a liquid state to a gel state as a result of ionic interaction circumstances at room temperature.

Production of nanoparticles using supercritical fluid technology

Traditional methods such as solvent extraction-evaporation, solvent diffusion, and organic phase separation need the use of organic solvents, which are hazardous to the environment as well as to human physiological systems. As a result, supercritical fluid technology is being investigated as an alternative to biodegradable microbes and nanoparticles, due to the fact that supercritical fluids are completely safe to the environment. Typically, a supercritical fluid is defined as a solvent at a temperature above its critical temperature, at which point the fluid's relationship to pressure is irrelevant for a single phase of the fluid. In part due to its mild critical conditions (Tc=31.1°C, Pc=73.8 bars), non-toxicity, non-flammability, and low cost, supercritical CO2 is the most widely used supercritical fluid. It is also the most widely distributed supercritical fluid. Among the most common processing techniques for supercritical fluids, supercritical solvent control (SAS) and rapid expansion of critical solution are two of the most often used (RESS). SAS makes use of a liquid solvent, such as methanol, that can be fully miscibly dissolved with a supercritical fluid in order to achieve its results (SC CO2). The extraction of the liquid solvent by means of a supercritical solvent result in immediate precipitation of the solution under the processing conditions, leading in the production of nanopa particles. Despite the presence of supercritical fluid, the solution remains insoluble.

Thote and Gupta (2005) Reported the application of the modified technique SAS for the development of nanoparticles for microencapsulative purposes of hydrophilic drugs dexamethasone phosphate drug. RESS differ from the SAS process by dissolving its solution in supercritical fluids (such as supercritical methanol) and then expanding the solution into a small lower pressure region, thereby significantly decreasing the solvent power of supercritical fluids and eventually precipitating the solution. The technology



is clean since it is generally devoid of solvents. For the polymeric nanoparticle's product, RESS and its modified method have been utilised. Specially developed equipment is required and is more costly for supercritical fluid technologies which are environment-friendly and suited for mass production.

Effect of nanoparticles characteristics on drug supply Size of the batch The main features of nanoparticles systems are particle size and size distribution. They define the distribution of nanopartments in vivo, their biological destiny, toxicity and targeted capabilities. In addition, the loading, release of drugs and stability of nanoparticles may also be affected. A large number of studies have shown that sub-micron-sized nanoparticles provide a variety of benefits as a drug system 24 over microparticles. Nanoparticles often have comparatively greater IP in comparison to microparticles and, owing to their limited size and relative mobility, are accessible for a broader variety of biological targets.

In a Caco-2 cell line25, Desai et al discovered that 100 nm nanoparticles exhibited a 2.5-fold higher uptake than 1 m microparticles and a 6-fold higher uptake than 10 m microparticles. In a follow-up research 26, nanoparticles entered all submucosal layers in a rat in situ intestinal loop model, while microparticles were mostly found in the epithelial lining.

Conclusion:

It was also shown that nanoparticles may penetrate the blood-brain barrier when hyper osmotic mannitol opens tight junctions, potentially allowing for long-term administration of therapeutic medicines for difficult-to-treat diseases like brain cancers. The blood brain barrier was crossed by two coated nanoparticles. Only submicrons can be effectively used in certain cell lines, but not bigger microparticles. The release of drugs is influenced by the size of the particle. Therefore, most related medicines would be on or near the particle surface, resulting in rapid drug release, for the smaller particles with a greater surface area. While, bigger particles have big nuclei that may encapsulate more medicines and slowly spread. The danger of particle aggregation during storage and transportation of nanoparticles is similarly higher for smaller particles. Nanoparticles with the lowest feasible size yet the highest stability are always a problem. The degradation of polymer by a particle size may also be influenced. For example, with increasing particle size in vitro 31, the rate of PLGA polymer breakdown increased. It was believed that degradation products produced by PLGA may spread readily in smaller particles, while degrading products are more likely to stay within the polymer matrix for a longer length of time to lead to autocatalytic material deterioration. Consequently, bigger particles were theorised to promote the rapid breakdown of polymers and the release of drugs. However, Panyam et colleagues produced PLGA particles of various sizes and observed that the rates of in-vitro polymer breakdown for different sizes of particles were not very different 32. Currently, photon-relation spectroscopy or dynamic light dispersion is the quickest and most common technique for measuring particle size. The viscosity of the medium must be understood to photone correlation spectroscopy and determines the particle diameter via brownian movement and light dispersing characteristics. 33. Scanning or transmission electron microscopy typically verifies the findings achieved by photorelation-spectroscopy (SEM or TEM).



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